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The uterotonic screening of calyx extracts of *Hibiscus sabdariffa* on estrogenized isolated Uterus of Wister strain albino rats

Ogundeko Timothy Olugbenga ^{1, *}, Osagie Ize Anuoluwapo ², Bitrus James ³, Okoye Nkiruka Philomena ¹, Ogbole Emmanuel Anebi ¹, Edugbe Adikpe Emmanuel ³, Abobarin Olufunmilayo Ibiyemi ⁴, Moritiwon Olusayo ^{4, 5}, Ramyil Mamzhi Seljul Crown ⁶ and Bassi Amos Paul ²

¹ Department of Pharmacology and Therapeutics, College of Medicine and Allied Health Sciences, Bingham University, Jos Campus Nigeria.

² Department of Community Medicine & Primary Health Care, College of Medicine and Allied Health Sciences, Bingham University, Jos Campus Nigeria.

³ Department of Obstetrics & Gynaecology, College of Medicine and Allied Health Sciences, Bingham University, Jos Campus Nigeria.

⁴ Department of Histopathology & Morbid Anatomy, College of Medicine and Allied Health Sciences, Bingham University, Jos Campus Nigeria.

⁵ Department of Pharmacology and Toxicology, University of Jos, Nigeria.

⁶ Department of Microbiology & Parasitology, College of Medicine and Allied Health Sciences, Bingham University, , Jos Campus Nigeria.

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Abstract

The claim of the use of orally consumed concentrated *Hibiscus sabdariffa* calyx preparation for the purpose of abortion in view of its widely uses as a beverage in the present period of economic hardship where it is a preferred beverage due to its affordability, naturalness and for its numerous medicinal properties has posed another element of concern to the female folks. This study was aimed to screen extracts of *H. sabdariffa* calyx of its acclaimed uterotonic activity. Contractile responses of estrogenized isolated rat uterus using various volumes (0.1, 0.2, 0.4, 0.6, 0.8, 1.0, 1.6 ml) were established with oxytocin $(1x10^{-4} \text{ g/ml})$, ergometrine $(1x10^{-4} \text{ g/ml})$, misoprostol $(1x10^{-4} \text{ g/ml})$, Acetylcholine $(1x10^{-3} \text{ g/ml})$ g/ml) and five different extracts of H. sabdariffa - ethanol (2.0x10⁻³ g/ml), methanol (2.0x10⁻³ g/ml), acetone (2.0x10⁻³ g/ml), cold water ($2.0x10^{-3}$ g/ml) and hot water ($2.0x10^{-3}$ g/ml). After these, nifedipine, acetylcholine and salbutamol were used to challenge the maximal responses of the isolated uterine tissue to oxytocin, acetylcholine and the cold aqueous extract of H. sabdariffa (CWEHs). The CWEHs was equally potent at inducing contraction activity compared to the standard drugs. This study further shows reduction in the amplitude of contraction produced by CWEHs in the presence of Promethazine $(1 \times 10^{-6} \text{ g/ml})$ and Nifedipine $(1 \times 10^{-6} \text{ g/ml})$. The blockade is more pronounced in promethazine compared to Nifedipine. Aqueous extract of Hibiscus sabdariffa calyx possess uterotonic activity on estrogenized isolated rat uterine muscle with mechanism of action related to histamine receptors and contraction of the myometrial cells as the was antagonized by promethazine and Nifedipine. This could be a justification for its alarming use as an abortifacient.

Keywords: Abortifacient; Uterotonic; Hibiscus sabdariffa; Caution; Beverage; Cold water

* Corresponding author: Ogundeko Timothy Olugbenga

Department of Pharmacology and Therapeutics, College of Medicine and Allied Health Sciences, Bingham University, Jos Campus Nigeria.

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1 Introduction

The uterus is a hollow organ that provides protective and nutritive support to the ovum from the moment it is fertilized until it grows into a well-developed fetus ready for parturition. Its middle muscular layer is called the myometrium, which is known for its rhythmic contractions which result in 'endometrial waves' in the nonpregnant uterus, Braxton Hicks contractions during pregnancy, and true labor towards the end of the third trimester [1]. Abnormal uterine contractility may underlie important clinical pathologies such as infertility, implantation failure, ectopic pregnancies, spontaneous miscarriages, preterm birth, and caesarean section [2]. Miscarriage, also known as spontaneous abortion and pregnancy loss, is the natural death of an embryo or fetus before it is able to survive independently [3]. In other words, abortion refers to the spontaneous or deliberate ending of a pregnancy before the fetus has evolved sufficiently to survive. Overall, abortion is the termination of pregnancy before 20 weeks of pregnancy [4]. The causes of abortion include embryonic factors including chromosomal abnormalities and abnormal evolution of the zygote, and maternal factors including uterine anatomical abnormalities, immunological factors, coagulation disorders, endocrine factors, infections, environmental factors, physical trauma and cervical failure [5]. Induced abortion is the deliberate termination of pregnancy using medical or surgical procedures before the embryo can survive. In Nigeria, abortion is only legal to save a woman's life. Recent estimates extrapolated from facility-based abortion complications indicate there were approximately 33 abortions per 1000 women aged 15-49 in 2012 [6]. Findings from gynaecological admissions at nine referral hospitals in Nigeria suggest that, although surgical abortion is still the primary method of abortion, the share of postabortion care (PAC) patients who report first using misoprostol is increasing [7]. Furthermore, among gynaecological admissions at a Nigerian teaching hospital in recent years, 7.4% were related to treatment of unsafe abortion, 17% of which ultimately resulted in maternal death [8]. Efforts to increase awareness of the availability of medication abortion drugs to more safely self-induce can help mitigate the toll of unsafe abortionrelated morbidity and mortality [9]. It has been shown that over 60% of unintended pregnancies among adolescents in secondary school will invariably end up in induced abortion [10]. Lamina et al, (2015) also reported the prevalence of unintended pregnancy as 35.9% while that of induced abortion was 33.5% also with the country's economic condition often cited as reasons women (15-48years) make use of abortion services for among women in South-Western Nigeria [11].

The foregoing are indicators that abortion in Nigeria is a public health concern as it affects both the young and the old both directly and indirectly, thus a social challenge. Herbal medicines are also emmenagogues (to avoid uterine flow) and oxytocic's (to stimulate uterine tightening, predominantly to induce labor) [12]. The use of herbal medicine has been on the increase in many developing countries [13]. These herbal remedies are used due to their cost effectiveness and ease of access [14]. The percentage of women depending on herbal medicine for their healthcare needs in developing countries is about 80% [15]; restricting its use by pregnant women has been a herculean task. Most pregnant women assume that nature-based medications do not lead to any drug interaction for fetal and maternal and therefore turn to self-medication with them, while they may cause certain fetal and maternal side effects or drug interactions [16]. Indigenous varieties of plants from various families are used by pregnant mothers in rural areas and low-income populations of sub-Saharan Africa for their uterotonic potential [17]. Despite the fact that majority of local herbal remedies used by sub-Saharan African pregnant women have never been botanically identified [18], several plants have been screened for their uterotonic activity using *in vitro* methods with positive results [19]. Report from a community visit showed that adolescent girls orally consume high concentration of *Hibiscus sabdariffa* calyces drink for abortion.

The genus *Hibiscus* (Malvaceae) includes more than 300 species of annual or perennial herbs, shrubs or trees [20] including *Hibiscus sabdariffa* (*Hs*) [21], which became domesticated and widely cultivated West Africa, cutting across both tropical and subtropical regions especially Sudan, Nigeria, Egypt as well as Mexico in the Americas [22, 23]. *Hs* is easy to grow in most well drained soils but can tolerate poor soils. In Sudan and Nigeria, the calyces are boiled with sugar to produce a drink known as "Karkade" or "Zoborodo" [24]. Both clinical and non- clinical studies revealed some medicinal properties of *H. sabdariffa*, including decreased systolic and diastolic blood pressures in those patients [25], anti-inflammatory activity [26] antioxidant [27], anti-diabetic [28], antinociceptive effect in a rat model [29], anti-microbial [30] e.tc

The claim of the use of orally consumed concentrated *Hibiscus sabdariffa* for the purpose of abortion in view of its widely used as a beverage (Zoborodo) in the present period of economic hardship where it is a preferred beverage due to its affordability, naturalness and for its numerous medicinal properties has posed another element of concern to the female folks thus urgent need for pharmacological screening for uterotonic activity as the aim of this study.

2 Material and methods

2.1 Study Site

This study was carried in the Pharmacology Laboratory, Department of Pharmacology and Therapeutics, College of Medicine and Allied Health Sciences, Bingham University, Jos, Nigeria.

2.2 Plant material procurement and preparation

Dried leaves of *Hibiscus sabdariffa* was purchased from Terminus Market, Jos North LGA, Plateau State, Nigeria.

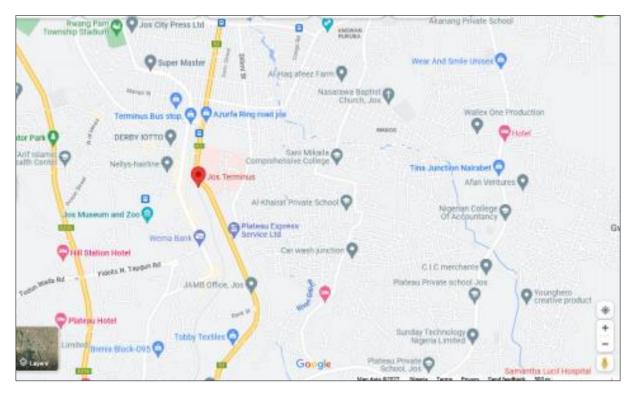


Figure 1 Google map of Terminus Market [31]



Figure 2 Dried calyx of H. sabdariffa [32]



Figure 3 Fresh plant of *H. sabdariffa* [33]

2.3 Plant material procurement and Preparation of *H. sabdariffa* Leaves Material

The procured dried calyces of *H. sabdariffa* were gently rinsed with clean water and dried under ambient light and temperature in a well-ventilated environment for 5 days, after which the dried leaves were ground into powder using mortar and pestle, The powder was then sieved into fine powder using size 80 Sieve. The Powder of plant material was stored in dark bottle container until required for use.

2.4 Extraction of *H. sabdariffa* Leaves Material

Five different extracts using different solvents (ethanol, methanol, acetone, cold water and hot water) were obtained. Powder sample of *H. sabdariffa* (100g) each was weighed and soaked in five different jars containing 700ml each of ethanol, methanol, acetone, cold water and hot water respectively stirred by way of extraction (maceration) for 72 hours. The extracts were filtered using Whitman No.1 filter paper while the crude extracts were then evaporated to dryness using a rotary evaporator and percentage yield calculated to obtain a powder whose yield was calculated. The five extracts eventually parked into dark bottles and stored in a refrigerator at 4°C according to the method of Idyu *et al.*, 2015 [34] and Builders *et., al* 2016 [35]. Until required for use.

Percentage yield =
$$\frac{\text{Mass of pure powder}}{\text{Mass of impure}} X 100$$

From the extraction of the extracts of *H. sabdariffa*, the yield was cold water (15.3%), Hot water (12.93%), ethanol (11.60%), methanol (8.20%) and acetone (4.1%) respectively.

2.5 Acute Toxicity Test

The LD₅₀ of the ethanol, methanol, acetone, cold water and hot water *H. sabdariffa* extracts were determined following Lorke's model [36]. Results from the phase 1 and 2 shows $10 \le x \le 5000$ for all the extracts.

2.6 Experimental Animals

Healthy non pregnant adult female rats (Wistar strain) rats between 10 and 12 weeks old and weighing 150–170 g were obtained from the Animal House Unit of the Department of Pharmacology and Therapeutics, College of Medicine and Allied Health Sciences, Bingham University, Jos, Nigeria. The animals were housed in plastic cages, allowed free access to pelleted animal feed and clean water *ad libitum* for 7 days for acclimatization in the Pharmacology Laboratory. This study was out carried in the Pharmacology Laboratory of the College of Medicine and Allied Health Sciences, Bingham University, Jos, Nigeria.

2.7 Standard Drugs and Physiological Salt solution Used

The following drugs were used for this study:

• Diethylstilbestrol injection (Kunj Pharma pvt, Ltd., India) was used to estrogenize the nongravid Wistar rats.

- Oxytocin injection (Rotex Medica), Ergometrine (Hameln. UK.) And Misoprostol (Marie Stop Int) were used as a standard uterotonic drug.
- Acetylcholine (Sigma Aldrich Germany).
- Atropine injection (Sigma Aldrich Germany) was used to block the muscarinic receptors of the uterine smooth muscles.
- Nifedipine (Krishat Pharma Ind. Nigeria) Inhibitor of Calcium channel blocker
- Indomethacin (Greenfield Pharm Ltd. China)
- Promethazine (JinLing Pharmaceutical, China)
- Freshly prepared De Jalon solution of the following composition: NaCl (9.00 g), KCl (0.42 g),
- CaCl2 (0.06 g), NaHCO3 (0.50 g) and glucose (0.50 g).

2.8 Animal Preparation/ Isolated Organ Preparation

The animals were estrogenized with 0.2 mg/kg diethylstilbestrol (DES) intraperitoneally (i.p). The drug was reconstituted with ethanol/water (1:1) solution prior to drug administration. Twenty-four hours later, the rats were sacrificed under chloroform anesthesia, had the uterine horns isolated devoid of excess fat and connective tissues, and cut into longitudinal strips. Then the uterine muscle strips were suspended with one end attached to a tissue holder in a 50 ml capacity tissue bath containing De Jalon's physiological solution. The other end of the strips was connected to an isotonic transducer and in turn connected to a 3 channel microdynanometer Power Lab (BD Instruments – ISO 9001:2000). The entire organ bath was maintained at 37°C and aerated with a mixture of 95% oxygen (O₂) in 5% carbon dioxide (CO₂). Having allowed the preparation to equilibrate for 30 minutes, tissue activity was monitored and observed via the microdynanometer recording paper before and after an intervention.

Contractile responses using various volumes (0.1, 0.2, 0.4, 0.6, 0.8, 1.0,1.6 ml) were obtained with oxytocin (1 x 10^{-4} g/ml), ergometrine (1 x 10^{-4} g/ml), misoprostol (1 x 10^{-4} g/ml), Acetylcholine ((1 x 10^{-3} g/ml) and five different extracts of *H. sabdariffa* - ethanol (2.0 x 10^{-3} g/ml), methanol (2.0 x 10^{-3} g/ml), acetone (2.0 x 10^{-3} g/ml), cold water (2.0 x 10^{-3} g/ml) and hot water (2.0 x 10^{-3} g/ml). After these, nifedipine, acetylcholine and salbutamol were used to challenge the maximal responses of the isolated uterine tissue to oxytocin, acetylcholine and the cold aqueous extract of *H. sabdariffa*.

The mechanism of *H. sabdariffa*-induced uterine contractions by way of cholinergic, histaminergic and oxytocinergic pathways were ascertained in the presence of antagonists by pre-treatment with atropine (1×10^{-6} g/ml), Promethazine (1×10^{-6} g/ml) used to block H₁ receptors, Nifedipine (1×10^{-6} g/ml) used to block the L-type calcium channel and Indomethacin (1×10^{-6} g/ml) used to inhibit cyclooxygenase (COX) enzymes of the uterine smooth muscles.

3 Result and Discussion

The present study shows that the hot water, ethanolic, methanolic and acetone extracts of *Hibiscus sabdariffa* most probably denatured the active ingredients in the plant sample, this suggests the reason for low percentage yield and zero contraction of the uterine muscle, in other words, administration of increasing doses of hot water $(2 \times 10^{-3} \text{ g/ml})$, ethanol $(2 \times 10^{-3} \text{ g/ml})$, methanol $(2 \times 10^{-3} \text{ g/ml})$, and acetone $(2 \times 10^{-3} \text{ g/ml})$ produced no contraction on the uterine smooth muscle and may be due to the fact that the active constituents in *Hibiscus sabdariffa* are more soluble in cold water. However, the cold-water extract has higher percentage yield (15.3%) and uterotonic activity at concentration of $2 \times 10^{-3} \text{ g/ml}$ on the isolated tissue.

Uterine smooth muscle often undergoes spontaneous rhythmic contractions that change in amplitude and frequency. Addition of increasing volumes of Misoprostol (1×10^{-4} g/ml), Oxytocin (1×10^{-4} g/ml), ergometrine (1×10^{-4} g/ml) dose dependently enhanced the contraction activity of the uterine muscles (amplitude and frequency), and produced a graded dose response tracing. Similarly, the CWEHs (2×10^{-3} g/ml) produced a dose dependent contraction of the rat uterine smooth muscle. The volume that gave the maximal response was 0.2ml (2×10^{-3} g/ml) – *Figure 4*. Administering greater than 0.2ml (2×10^{-3} g/ml) of the CWEHs is not likely to produce a more forceful contraction. One reason for this may be that the contractile mechanism is already running in a saturated manner. The responses of CWEHs were significant compared to the normal uterine contraction but the standard drugs were more potent especially oxytocin. The concentration-response curve gave sigmoidal with EC₅₀ for CWEHs (3.2×10^{-5} g/ml), Oxytocin (1.3×10^{-4} g/ml), Ergometrine (4×10^{-5} g/ml), Misoprostol (1.6×10^{-4} g/ml), and Acetylcholine (3.2×10^{-6} g/ml). This study further shows reduction in the amplitude of contraction produced by CWEHs in the presence of Promethazine (1×10^{-6} g/ml) and Nifedipine (1×10^{-6} g/ml) – *Figure 5*. The blockade is more pronounced in promethazine compared to Nifedipine.

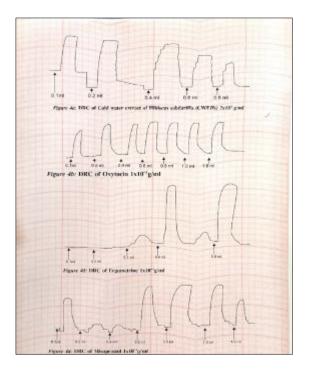


Figure 4 Dose- response curve (DRC) of CWEHs and standard drugs

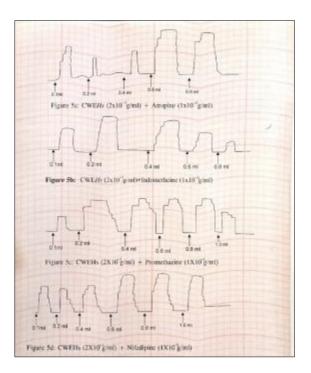


Figure 5 DRC of CWEHs in the presence of antagonists

Uterine contractile activity is regulated by the increase in intracellular Ca^{2+} concentration in the myometrial cells. Voltage-gated Ca^{2+} channels (VGCCs) mediate the Ca^{2+} influx in response to membrane depolarization and regulate intracellular processes such as contraction [37, 38], 39]. Also, Histamine receptors (H₁ and H₂) are present in the uterus and the predominant response of histamine in this tissue is contraction (H₁ activity) [40]. The antagonism activity of CWEHs in the presence of Promethazine and Nifedipine suggests the likely mechanism of action to be via H₁ receptors and by calcium mobilization in the myometrium. Uterotonic activity of the cold water or cold aqueous extract of *Hibiscus sabdariffa*, even though at low volume as highlighted in this study is in consonance with the report by Fofie and Baffoe,

2010, on the prevalence of uterine rupture experienced by pregnant women as a result of the use of herbal preparations for inducing labour [41] thus, a concern for the pharmacology, public health, gynaecology, and other field of medicine community. Mode of preparation before consumption of the plant product should be looked into, women in their third trimester should apply caution in taking beverages especially such prepared via soaking *H. sabdariffa* cold water. While the results presented convincingly show that the cold aqueous extract of *H. sabdariffa* induces contractions in uterine smooth muscle tissue in vitro, further information is warranted in determining whether if the same results can be seen *in vivo* [42] when the herbals are ingested as water, spirit, or oil-based medicines [43].

4 Conclusion

Aqueous extract of *Hibiscus sabdariffa* calyx possess uterotonic activity on estrogenized isolated rat uterine muscle with mechanism of action related to histamine receptors and contraction of the myometrial cells as the was antagonized by promethazine and Nifedipine. This could be a justification for the alarming use as an abortifacient.

Compliance with ethical standards

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Disclosure of conflict of interest

All the authors of hereby declare no conflicts of interest regarding this study.

Statement of ethical approval

Ogundeko T.O, Okoye NP among the authors and Kamoh L are licensed to handle laboratory animals thus, standard protocols involving the use of laboratory animals were strictly adhered to.

References

- [1] McEvoy A, Sabir S. Physiology, Pregnancy Contractions. StatPearls Publishing; 2022 Jan. [Updated 2021 Sep 20]. [cited 2022 Aug 17] Available from https://www.ncbi.nlm.nih.gov/books/NBK532927/
- [2] Birch Petersen K, Pedersen NG, Pedersen AT, Lauritsen MP, la Cour Freiesleben N. Mono-ovulation in women with polycystic ovary syndrome: a clinical review on ovulation induction. Reprod Biomed Online. 2016;32(6):563–83. doi: 10.1016/j.rbmo.2016.03.006.
- [3] Robinson GE. Pregnancy loss: Best Practice & Research. Clinical Obstetrics & Gynaecology. 2014;28(1):169–178.
- [4] Lohr PA, Hayes JL, Gemzell-Danielsson K. Surgical versus medical methods for second trimester induced abortion. Cochrane Database Syst Rev 2008. [cited 2022 Jul 15]
- [5] Edmonds DK, Lees C, Bourne TH. Dewhurst's textbook of obstetrics & gynaecology. Wiley Online Library; 2007.
 [cited 2022 Jul 17]
- [6] Bankole A, Adewole IF, Hussain R, Awolude O, Singh S, Akinyemi JO. The Incidence of Abortion in Nigeria. Int Perspect Sex Reprod Health. 2015 Dec;41(4):170-81. doi: 10.1363/4117015. PMID: 26871725; PMCID: PMC4970740.
- [7] Bello FA, Fawole B, Oluborode B, Awowole I, Irinyenikan T, Awonuga D, Loto O, Fabamwo A, Guest P, Bela Ganatra B. Trends in misoprostol use and abortion complications: a cross-sectional study from nine referral hospitals in Nigeria. PLoS One2018;13: e0209415.
- [8] Akinlusi FM, Rabiu KA, Adewunmi AA, Imosemi OD, Ottun TA, Badmus SA. Complicated unsafe abortion in a Nigerian teaching hospital: pattern of morbidity and mortality. Jol of Obst and Gynaecol . 2018 Oct 3;38(7):961-6.
- [9] Bell SO, Omoluabi E, OlaOlorun F, Shankar M, Mooreau C. Inequities in the incidence and safety of abortion in Nigeria. BMJ Global Health 2020;5: e001814.

- [10] Ayuba II, Gani O. Outcome of teenage pregnancy in the Niger Delta of Nigeria. Ethiop J Health Sci. 2012; 22: 45-50.
- [11] Lamina MA. Prevalence and Determinants of Unintended Pregnancy Among Women in South-Western Nigeria. Ghana Med J. 2015 Sep;49(3):187-94. doi: 10.4314/gmj.v49i3.10. PMID: 26693195; PMCID: PMC4676590.
- [12] Feroche T. Evaluation of abortifacient efficacy of Rumex steudelli (Tult) root traditionally used medicinal plant in South West Ethiopia. J. Pharmacog. Phytom. Res., 2015;4(4): 221-223.
- [13] Rouhi-Boroujeni H, Heidarian E, Rouhi-Boroujeni H, Khoddami M, Gharipour M, Rafieian Kopaei M. Use of lipidlowering medicinal herbs during pregnancy: A systemic review on safety and dosage. ARYA Atheroscler. 2017: 13(3): 135-155
- [14] John, LJ, Shantakumari N. Herbal medicines use during pregnancy: A review from the Middle East. Oman Med J. 2015; 30(4): 229-236.
- [15] Ameade, EPK, Zakaria AP, Abubakar L, Sandow R. Herbal medicine use before and during pregnancy a study in Northern Ghana. Int. J. Complement Alt. Med. 2018;11(4): 235-242.
- [16] Taghikhani M, Nasri H, Asgari A, Afrough H, Namjoo A, Ansari Samani R, et al. The renal toxicity of hydroalcoholic extract of Stachys lavandulifolia Vahl in Wistar rats. Life Sci J-Acta Zhengzhou Univ Overseas Ed. 2012; 9(4):3025e31.
- [17] Tripathi V, Stanton C, Anderson FWJ. Traditional preparations used as uterotonics in Sub-Saharan Africa and their pharmacologic effects. International Journal of Gynecology & Obstetrics. 2012; 120 (1): 16–22.
- [18] El Hajj M, Holst L. Herbal Medicine Use During Pregnancy: A Review of the Literature with a Special Focus on Sub-Saharan Africa. Front Pharmacol. 2020 Jun 9;11:866. doi: 10.3389/fphar.2020.00866. PMID: 32581815; PMCID: PMC7296102.
- [19] Bwalya P, Factors Influencing Use of Traditional Medicine to Precipitate Labour by Antenatal Mothers in Mpika District, University of Zambia, Lusaka, Zambia. 2010. [cited 2022 Jun 27], Available from http://dspace.unza.zm/handle/123456789/1940.
- [20] Wang ML, Morris B, Tonnis B, Davis J, Pederson GA. Assessment of oil content and fatty acid composition variability in two economically important Hibiscus species. Journal of Agricultural and Food Chemistry. 2012; 60 (26): 6620-6626.
- [21] The Plant List. A working list of all species. 2010 [cited 2022 Jul 27] Available from http://www.theplantlist.org/tpl/record/kew-2850461.
- [22] Morton JF. Fruits of warm climates. Florida Flair Books (1987). Google Scholar
- [23] Ibrahim KG, Kangiwa NS. A Review of the Phytochemistry and BiologicalActivities of *Hibiscus sabdariffa* (Zobo) Plant. Annals of clinical and experimental medicine. 2020; 1(1): 1-10.
- [24] D. Gibbon, A. Pain. Crops of the drier regions of the tropics. (1st ed.), English Language Book Society, Longman, England (1985).
- [25] Mozaffari-Khosravi H, Ahadi Z, Barzegar K. The effect of green tea and sour tea on blood pressure of patients with type 2 diabetes: a randomized clinical trial. Journal of Dietary Supplements. 2013; 10 (2): 105-115.
- [26] Beltran-Debon R, Alonso-Villaverde C, Aragones G, Rodriguez-Medina I, Rull A, Micol V, et al. The aqueous extract of *Hibiscus sabdariffa* calices modulates the production of monocyte chemoattractant protein-1 in humans. Phytomedicine. 2010; 17 (3–4): 186-191.
- [27] Frank T, Netzel G, Kammerer DR, Carle R, Kler A, Kriesl E, et al. Consumption of *Hibiscus sabdariffa* L. aqueous extract and its impact on systemic antioxidant potential in healthy subjects. Journal of the Science of Food and Agriculture. 2012; 92 (10): 2207-2218.
- [28] Peng CH, Chyau CC, Chan KC, Chan TH, Wang CJ, Huang CN. *Hibiscus sabdariffa* polyphenolic extract inhibits hyperglycemia, hyperlipidemia, and glycation-oxidative stress while improving insulin resistance. Journal of Agricultural and Food Chemistry. 2011; 59 (18): 9901-9909.
- [29] Ali MK, Ashraf A, Biswas NN, Karmakar UK, Afroz S. Antinociceptive, anti-inflammatory and antidiarrheal activities of ethanolic calyx extract of *Hibiscus sabdariffa* Linn. (Malvaceae) in mice. Zhong Xi Yi Jie He Xue Bao. 2011; 9 (6): 626-631.

- [30] Nwaiwu NE, Mshelia F, Raufu IA. Antimicrobial activities of crude extract of Moringa Oleifera, *Hibiscus sabdariffa* and Hibiscus esculentus seeds against some enterobacteria. Journal of Applied Phytotechnology in Environmental Sanitation. 2012;1(1): 11-16.
- [31] Google map of Terminus market, Jos, Nigeria. [2022 Jul 10], Available from https://www.google.com/maps/place/Jos+Terminus/@9.9165483,8.8903481,15z/data=!4m5!3m4!1s0x0:0xa 5eaa035e99c8605!8m2!3d9.9165569!4d8.8904038?hl=en-GB
- [32] Dried leaves of *Hibiscus sabdariffa* [cited 2022 Jul 10], Available from https://ocdn.eu/pulscmstransforms/1/lAQktkpTURBXy9lOTFlMTc0MWU0NWM1MzcwOWUyMDUxOTJjYmI0ZjQyYS5qcGeSlQMAP80E AM0CQJMFzQSwzQJ2
- [33] Fresh plant of *Hibiscus sabdariffa*. [cited 2022 Jul 10], Available from https://www.nairaland.com/attachments/3684626_1312493410541541146305615422229118345340957n_jpgc4ea91fd5b5716e3be44cf0f0ff58690
- [34] Idyu II, Deshi EF, Idyu VC, Ogundeko TO. The Anti-diarrhoeal Effect of Ethanolic-Bark Extract of Sterculia setigera in Mice. Asian J. Sci. Tech. 2015; 6(5): 1397-1400.
- [35] Builders MI, Builders PF, Ogundeko TG. Anti-ulcer activity of the stem bark of African locust bean tree in rats. Int. J. Phytotherapy Research. 2016; 6 (4). 11-19.
- [36] Erhirhie EO, Ihekwereme CP, Ilodigwe EE. Advances in acute toxicity testing: strengths, weaknesses and regulatory acceptance. Interdiscip Toxicol. 2018 May;11(1):5-12.
- [37] Gáspár R, Hajagos-Tóth J. Calcium channel blockers as tocolytics: principles of their actions, adverse effects and therapeutic combinations. Pharmaceuticals (Basel). 2013 May 23;6(6):689-99. doi: 10.3390/ph6060689. PMID: 24276256; PMCID: PMC3816733
- [38] Catterall WA., Perez-Reyes E, Snutch TP, Striessnig J. International Union of Pharmacology. XLVIII. Nomenclature and structure-function relationships of voltage-gated calcium channels. Pharmacol. Rev. 2005; 57:411–425. doi: 10.1124/pr.57.4.5.
- [39] Wray S, Jones K, Kupittayanant S, Li Y, Matthew A, Monir-Bishty E, Noble K, Pierce SJ, Quenby S, Shmygol AV. Calcium signaling and uterine contractility. J. Soc. Gynecol. Investig. 2003; 10:252–264. doi: 10.1016/S1071-5576(03)00089-3.
- [40] Martinez-Mir I, Herrero J, Estan L, Morales-Olivas FJ, Rubio E. Effect of histamine on the longitudinal and circular muscle of the oestrogen dominated rat uterus. Agents and Actions.1993; 39 (1-2): 1–5, 1993
- [41] Fofie C, Baffoe P. A two-year review of uterine rupture in a regional hospital. Ghana Medical Journal. 2010; 44 (3): 98–102.
- [42] Lin J. Applications and limitations of interspecies scaling and in vitro extrapolation in pharmacokinetics. Drug Metabolism and Disposition 1998; 26:1202-1212.
- [43] Weed S. Planting the Future: Saving Our Medicinal Herbs, 2000 books.google.com.